



FIGURE 5. FREQUENCY OF RAT AND MOUSE PRINTS ON PAPERS FROM TRACKING TUNNELS SET IN TREATMENT AND NON-TREATMENT SITES AT KARIOI RAHUI, OHAKUNE. THREE EXPERIMENTAL TRIALS (RUNS) OF VARYING LENGTH WERE SET, AT TWO WEEK INTERVALS, USING 15 TUNNELS IN EACH SITE. (NOTE: THREE NIGHTS TRACKING FOR RUN 1, TWO NIGHTS TRACKING FOR RUNS 2 AND 3.)

5. Discussion

This study may be confounded by the presence of rats and mice in both the treatment and non-treatment sites, although rat density was apparently low in the treatment site. In another study, mouse numbers have been observed to increase (following initial declines) after poisoning, regardless of the toxin or poisoning method (aerial or bait stations) (Innes *et al.* 1995). In that work, the increase in mouse numbers was greater in sites where ship rats had been most

effectively removed, implying that mice may survive better when inter-specific competition and predation from rats is reduced. This might explain our observation of high mouse presence in the treatment site at Karioi. Therefore, while the removal of possums and rats would be expected to have a benefit to invertebrate populations, predation pressure on invertebrates may be maintained by an increased abundance of mice, reducing the actual benefits of the poisoning operation to invertebrates.

Our invertebrates capture data tended to support the contention that rodents were still having an effect as predators in the treatment area. Both pitfall and malaise trap samples showed significant differences in the numbers of individuals in taxa which were collected in the treatment and non-treatment sites. However, the trends are not strongly defined, with some taxa occurring more frequently in the treatment area, and others more frequently in the non-treatment area. Araneae and Coleoptera >5 mm were scored more frequently in the non-treatment site in both pitfall and malaise samples, and adult Lepidoptera occurred more frequently in the treatment samples of both pitfall and malaise traps. Other taxa were recorded less consistently in the two trap types, and no real trend in terms of invertebrate abundance in one site or the other can be determined.

This situation may be the result of mice numbers not dropping because the poison bait stations were spaced too far apart relative to mouse home range size. Thus mice may still persist at densities sufficient to remain as significant predators of invertebrates. This scenario is supported by the ambivalent results from the statistical comparisons where significant differences occurred, but not necessarily with higher numbers in the treatment area, as one might expect if rodent predation was reduced by poisoning.

In order to establish an experiment in which the effects on invertebrates of rodent removal can be tested, rodent populations must be further reduced, and a poisoning regime which is effective on both mice and rats must be used. However, for research to be generally applicable, the poisoning programme and resulting changes in pest mammal abundance must represent the results of a typical poison operation. Creating an atypical situation, while showing the theoretical advantages for invertebrates if possums and all rodents were removed, would not tell us the actual benefits of mammal control operations in New Zealand.

The presence of rodents, especially mice, in the treatment site, and the small scale of the study, mean that we have not recorded data from which it is possible to gather evidence of the effect of rodent and possum removal on the abundance of invertebrates. Hence the pilot study was a trial of the methodology, and enables refinement of sampling methods for future work.

6. Recommendations for future research

Comparisons of invertebrate abundance and diversity between randomly selected treatment and non-treatment areas needs to be carried out in advance of the application of toxin—probably over two seasons. Further comparisons of invertebrate abundance and diversity between the treatment and non-treatment sites would be required for at least two years during toxin application (via poison bait stations which are accessible to all rodents as well as possums) followed by at least two years monitoring after the application of toxin. This will enable any changes in invertebrate numbers to be attributed to the reduction of pest mammals. The investigation of benefits of pest mammal control operations to invertebrates is therefore a long-term research project, requiring six years of work in the field.

In particular, it seems that longer sampling periods (perhaps six weeks) are required in beech forest to adequately sample large-bodied invertebrates using pitfall or malaise traps. Techniques such as “fences” which increase the catch area of pitfall traps or large diameter traps should be used. These have been shown to increase the proportion of larger beetles which are collected (Southwood, 1978).

New sampling methods may be required, especially if particular invertebrate species are found to be preferred in the diet of rodents and possums. Standardised techniques which involve manual searching should be used. The data yielded would be catch rates per unit effort. Pitfall and malaise trapping would need to be verified as methods which actually capture the invertebrate species which were found in rodent and possum diets.

Mammal species must be monitored using standard techniques throughout the invertebrate study, so changes in invertebrate density can be correlated with changes in mammal numbers. In addition, diet studies of rodents and possums will be required to ensure that the invertebrates sampled were actually prey items.

In this study invertebrates were generally not sorted to species level. Thus differences in species diversity may not have been apparent between samples because so many taxa were lumped together. Future work needs to compare changes in diversity (species composition) between sites as well as biomass.

7. References

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8. Acknowledgements

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Appendix 1

MALAISE TRAP SAMPLES, ORIGINAL FREQUENCY SCORES

Taxa larger than 5mm	month total	all taxa	NT	T
Litho	0	71	1	82
Aran	4	4	5	9
Opi	4	4	5	9
Pseud	8	45	5	50
Collemb	5	36	4	40
Cellatob	2	2	2	4
isoplec	0	0	0	0
Gy tu	5	5	5	10
Hemidein	1	1	1	2
Crick	0	0	0	0
Staph	3	34	3	37
Coleopt lrg	0	0	0	0
Coleopt sm	2	2	2	4
Coleopt larv	0	0	0	0
SS	1	1	1	2
Odont	6	6	6	12
Cerambyc	0	0	0	0
Chrys	2	2	2	4
Lepid	0	0	0	0
Lepid larv	4	4	4	8
Calliph	1	1	1	2
Tach	9	9	9	18
Hoverf	3	3	3	6
DIPT:sm d	1	1	1	2
DIPT:lrg	7	7	7	14
DIPT:lrg bow	3	3	3	6
DIPT:smi	4	4	4	8
DIPT:hairy	2	2	2	4
DIPT:stripy	5	5	5	10
DIPT:yellow abd	1	1	1	2
DIPT:lrg y dros	0	0	0	0
DIPT:smi byw	5	5	5	10
DIPT:y Drosop	19	19	19	38
DIPT:ybw	0	0	0	0
Hymenop	3	3	3	6
Ichnumon	1	1	1	2
Apidae	3	3	3	6
Pomp	3	3	3	6
Termite?	5	5	5	10
Bee?	3	3	3	6
Vespu	11	11	11	22
Hemipt	27	27	27	54
Psocopt	12	12	12	24
like Psocopt	5	5	5	10
Mayfly	0	0	0	0
Lacewing	2	2	2	4
Cranefly	80	80	80	160
Cranefly smi	44	44	44	88
Robberfly	15	15	15	30
Robberfly sml	7	7	7	14

Taxa	NT	T
Aranaea	71	82
Opilioniidae	4	9
Blattodea	4	9
Orthoptera	8	10
Staphylinidae	34	37
Coleoptera - large (>5mm)	2	4
Larvae (Coleopt. & Lepidopt.)	4	8
Lepidoptera (adult)	0	0
Hemiptera	1	2
Ephemeroptera	3	6
Neuroptera	2	4
Tipulidae	0	0

Taxa	NT	T
Lithobiomorpha	0	0
Araneae	6	9
Opilioniidae	1	1
Pseudoscorpionidae	1	1
Collembola	13	15
Cellatoblatella vulgaris	1	1
Isopleuron (gold stripe)	1	1
Gymnoleptus luanii	1	1
Hemideina	1	1
Crick	0	0
Staphylinidae	3	3
Coleoptera - large (>5mm)	7	7
Coleoptera - small (<5mm)	12	12
Coleoptera larvae	3	3
Saphobius squam.	1	1
Odontria	1	1
Cerambycidae	2	2
Brown Chrysomelidae?	15	15
Lepidoptera	28	28
Lepidoptera larvae	12	12
Calliphoridae	2	2
Tachinidae	7	7
Hoverfly?	3	3
DIPT: assorted small dark	9	9
DIPT: various large, slender (>5mm)	34	34
DIPT: Large brown, orange wing	4	4
DIPT: various small (<5mm)	22	22
DIPT: brown hairy & large hairy	10	10
DIPT: stripy housefly?	13	13
DIPT: yellow abdomen	8	8
DIPT: large, like yellow thorax Drosophila	1	1
DIPT: Small brown, yellow wing	4	4
DIPT: Yellow thorax drosophila	15	15
DIPT: yellow body, spotted wing	6	6
Hymenoptera - small	11	11
Ichneumonidae (large)	5	5
Apidae	3	3
Pompilidae	2	2
Winged termite/ant?	2	2
?Bee?	10	10
Vespa vulgaris	18	18
Hemiptera	3	3
Psocoptera?	20	20
? like Psocoptera?	1	1
Mayfly	1	1
Lacewing	4	4
Cranefly	4	4
Small Cranefly (<10mm)	75	75
Robberfly	1	1
Robberfly - small	7	7

Appendix 2

PITFALL TRAP SAMPLES, ORIGINAL FREQUENCY SCORES

Taxa	T	NT
Endodontidae	4	3
Amphipoda	116	109
Isopoda	19	14
Diploida & Chilopoda	67	59
Araneae	316	342
Opiliones	22	18
Blattodea	3	52
Orthoptera	52	21
Larvae (Lep., Dipt. & Coleopt.)	44	44
Carabidae	15	15
Staphylinidae	4	0
Coleoptera>5mm (ex Carabids)	0	4
Neuroptera	1	1
Lepidoptera (adult)	1	0
TOTAL	1684	781

Taxa larger than 5mm, month totals for pitfall trap samples. T=treatment, NT=non-treatment.

Taxa	T	NT
Endod	1	0
Otoconc	1	1
Plan	1	1
worms	4	2
Amph	116	109
Sphar	4	2
Phil	2	13
SlaB	3	2
Diplod	3	2
Dimer	1	2
Jul	55	50
Icosid	7	2
Litho	0	1
Corm	1	2
Aran	316	342
Acar	197	182
Opil	22	18
Psued	11	17
Collem	507	602
Cellato	3	1
Rhap	0	10
Zeland	0	1
Weta	45	76
Coleop	110	92
Col. lar	1	8
Car1	28	12
Car2	8	5
Mep1	8	1
Stap	11	49
Qued	4	4
Odont	3	4
SS	298	105
Cucu	0	1
choc	1	2
Lep larv	16	61
Dipt?	806	83
Dipt larv	4	0
Apt dipt	55	1
Callip	84	2
Tach	7	0
Hymen	97	83
Form	48	186
Ich	2	3
Pomp	3	1
Hemip	7	1
Lace	0	1
Snail	2	2
thing	1	0
Rhyp	1	0
Lep	1	0
TOTAL	2723	2111

All taxa, month totals for pitfall trap samples. T=treatment, NT=non-treatment

Taxa	T	NT
Endodontidae	1	0
Otoconcha dim.	1	0
Planeria	1	0
worms	4	2
Amphipoda	74	31
Sphaerillo	2	6
Philoscia	3	7
Slater B	2	1
Diploida - unknown	6	0
Dimerogonus	3	1
Julida	1	27
Icosidesmus	5	0
Lithobiomorpha?	0	1
Cormocephalus rubiceps?	0	1
Araneae	169	176
Acarinae	107	92
Opiliones	16	10
Psuedoscorpionidae	9	7
Collembola	255	241
Cellatoblatta vulg.	2	0
Rhaphid/Gymnoplec. tuarti	4	3
Zelandrosandrus	1	1
Weta nymph	21	37
Coleoptera - small unident	79	27
Coleoptera larvae	0	4
Carabid 1	17	4
Carabid 2	4	3
Mecyclothorax plac.	6	1
Staphylinidae	9	4
Quedius tinctellus	4	0
Odontria	4	0
Saphobius sqam.	1	0
Cucujidae?	0	0
choc chip/wheel arch beetle	2	0
Lepidopt larvae	12	9
Diptera - small unident	150	20
Dipteran larvae	2	1
Wingless fly	11	0
Calliphoridae	34	2
Tachinid fly	4	0
Hymenoptera - small unident.	51	58
Formicidae (incl Huberia browni)	29	53
Ichneumonidae	1	2
Pompilidae	1	0
Hemiptera	1	1
Lacewing	2	0
Snail	2	2
"thing" (Mike's drawing)	1	0
Rhyphodes	1	0
Lepidoptera	1	0
TOTAL	1339	872

All taxa, two week samples for pitfall traps. T=treatment, NT=non-treatment