

# INT2022-05 Determining the resilience of Fiordland corals to fisheries impacts



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**Associate Professor Peter Ritchie**  
**Ms Miriam Pierotti**  
**Dr Alice Rogers**



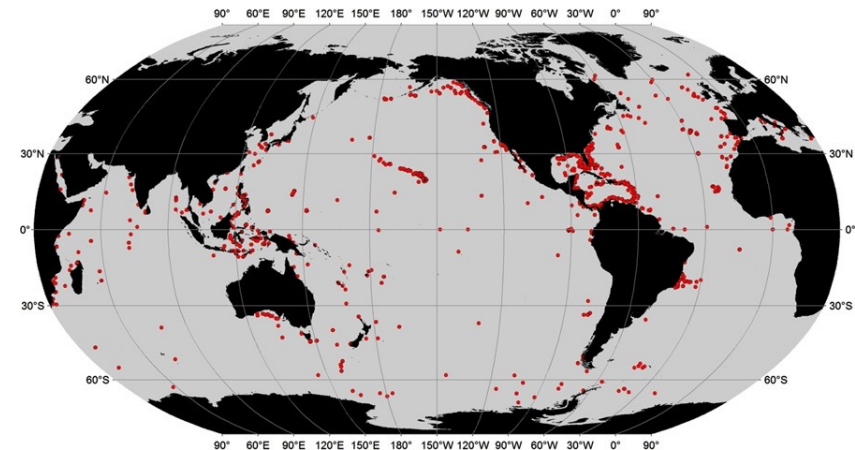
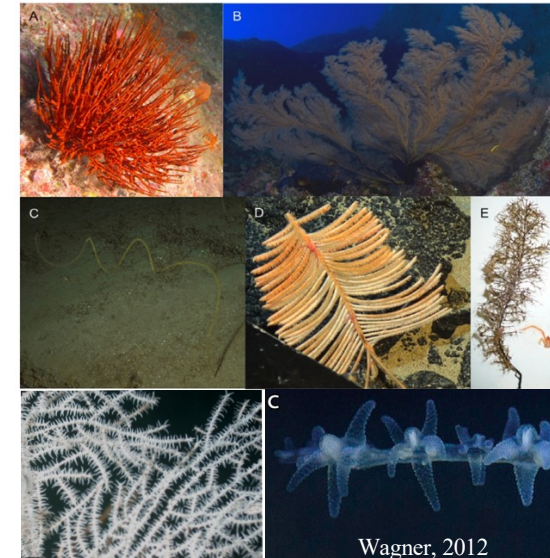
# Broad project objectives:

1. Improve our understanding of the distribution of Fiordland black corals inside and outside of fished areas and ascertain the extent of overlap between fishing activity and coral habitat.
2. Increase understanding of the ecology and impacts of fishing on protected corals in Fiordland, including the black coral *Antipathella fiordensis* and stylasterid corals.
3. Use varied approaches (modelling, SCUBA and remotely operated vehicle ('ROV') surveys, pre-existing data) to inform our understanding of protected coral resilience to fishing impacts and threats in Fiordland, which can then be applied to these taxa in a wider context
4. Determine patterns of genetic diversity and likely routes of connectivity within and between Fiords.

# Antipatharians

## Biology and Ecology

- Subphylum Anthozoan; class: Hexacorallia
- Colonial organisms with a wide range of morphology
- Ahermatypic
- 75% below 50 m
- Found in very low light environments below the photic zone



Global Antipatharians distribution  
Wagner, 2012



# Antipatharians

## Biology and Ecology

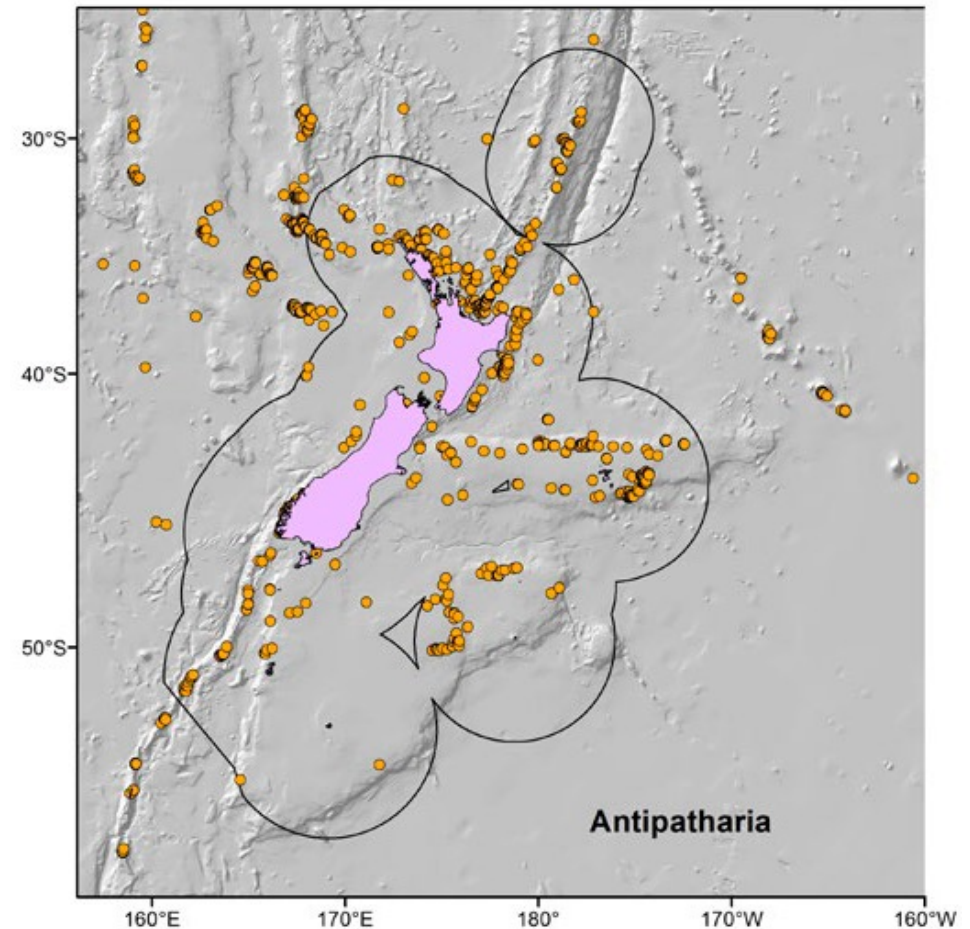
- Slow-grow rates / Longevity
- Habitat engineers
- Support sea-floor associated biodiversity and productivity
- Reproduction through both sexual and asexual processes. In general, polyps and colonies are gonochoric





# Black corals in New Zealand

- Predominantly deep-water group
- Around 60 described species found in New Zealand and another 20+ undescribed
- 1 'endemic' genus to the Fiordland region, *Antipathella fiordensis*.



Recorded Distribution of Antipatharia around NZ

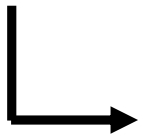


*Antipathella fiordensis*



# Current knowledge

- Early reproductive and population genetic studies with allozymes (Grange, 1990; Miller, 1997)
- Growth/ultrastructure (Goldberg, 1991)
- Relationships with other mutualistic species (Parker et al., 1997) – *Astrobrachion constrictum*
- Distribution limits in relation to salinity (Jiang et al., 2015)
- Age (Hitt et al. 2020)



Less information on how resilient they are to different forms of disturbance and their recovery potential



# Antipathella fiordensis

## Fiordland Marine Management Act (FMMA 2005)



## Recreational Fishing Rules FIORDLAND



Effective from: November 2022 (subject to change without notice).

### Doubtful Sound (Patea) fiord complex blue cod restricted area

Within the internal waters of Doubtful, Thompson and Bradshaw Sounds the daily take and possession limit is one blue cod per person with no accumulation.



# Study location

## DOUBTFUL SOUND

40.4 km long, the deepest of all fiords (434 m)

Spit in 3 distinct arms - Hall Arm, Crooked Arm, First Arm and 2 outer Sounds- Bradshaw and Thompson Sounds

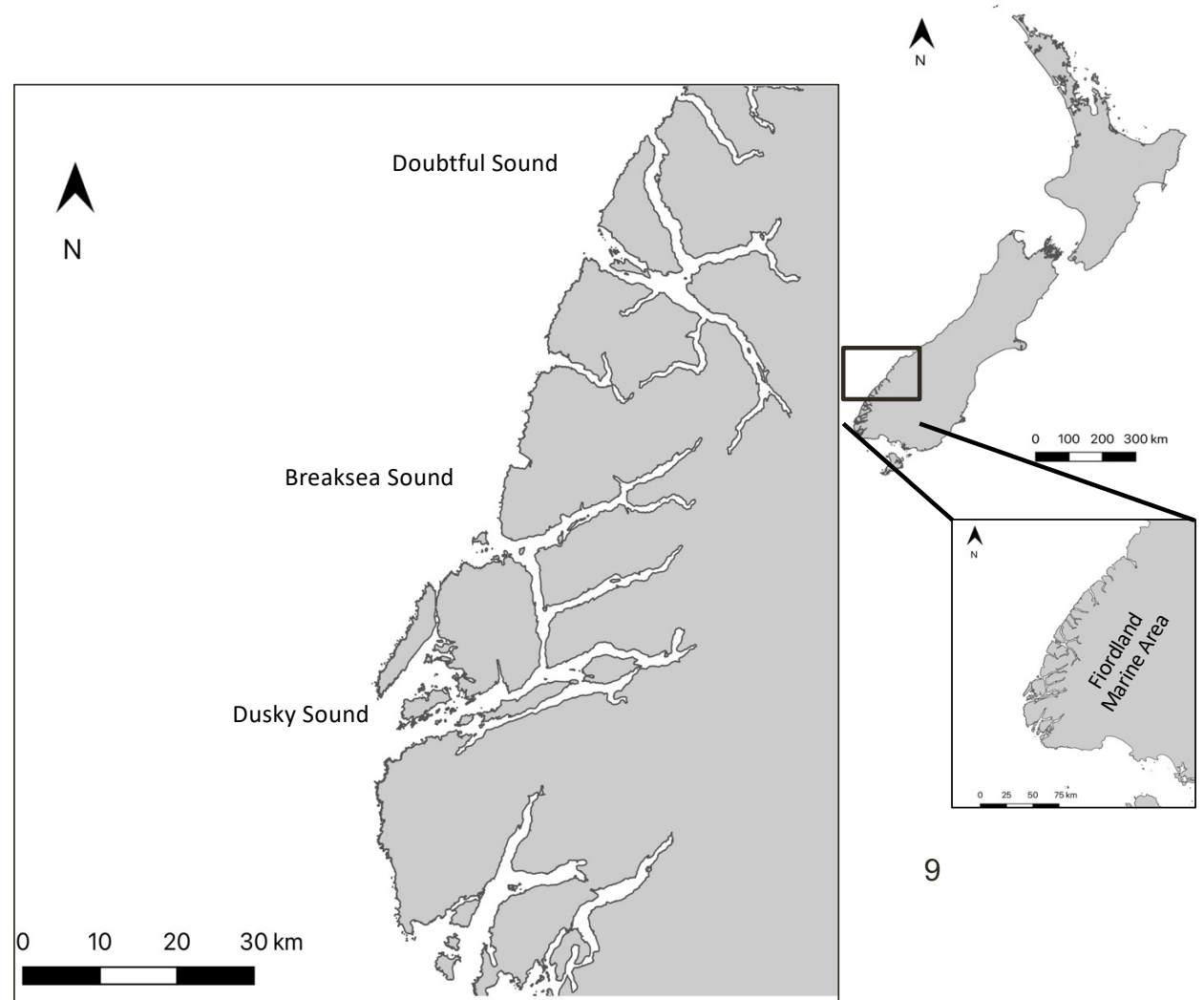
Manapouri Hydropower station

## BREAKSEA-DUSKY SOUNDS SYSTEM

Biggest fiord complex with Acheron passage (15km) connecting the two Sounds

Breaksea (33km) split in 2 arms – Vancouver Arm and Broughton Arm

Dusky is the longest and most extensive fiord (43.9 km) split into 2 main channels



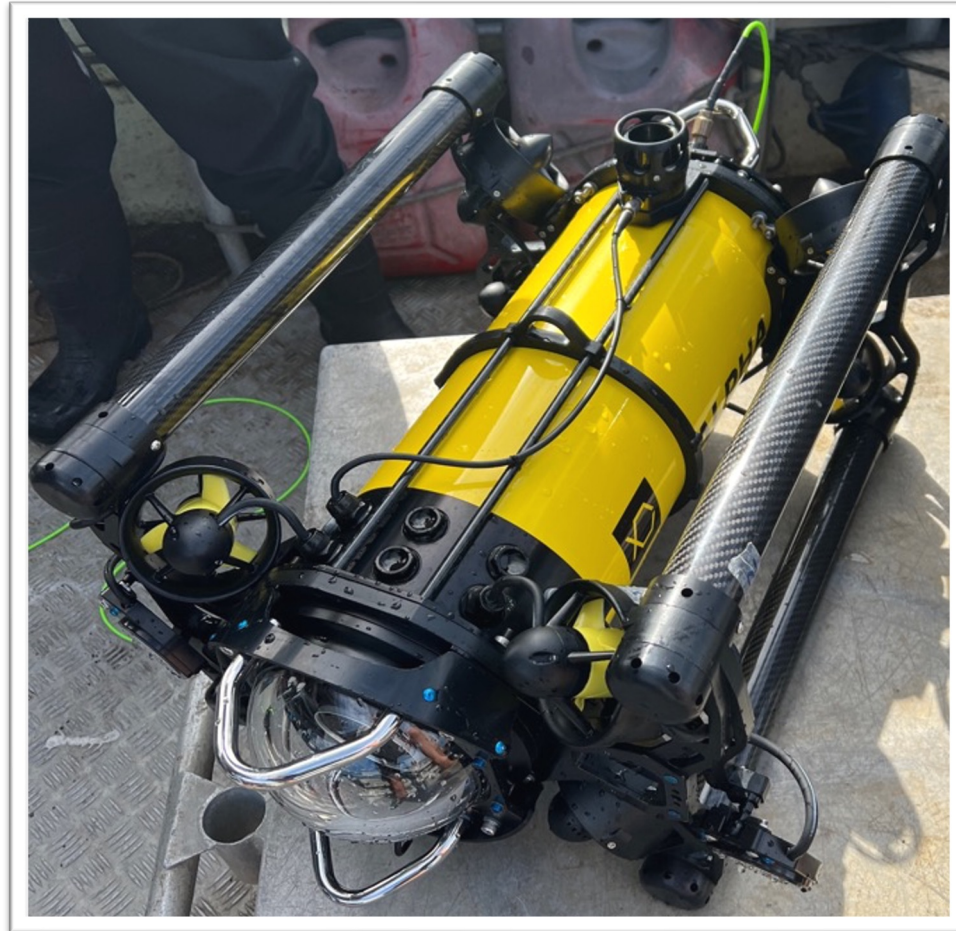
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## Investigate the spatial distribution and population size-structure

- Assessing the spatial and size frequency distribution along a vertical and horizontal gradient to quantify the abundance and size structure within and across different fiords
- Characterise distribution patterns across fiords and with depth
- Identify environmental variables that best predict the distribution



# Methods

## FIELD SAMPLING

## Abundance and Size

SCUBA diving

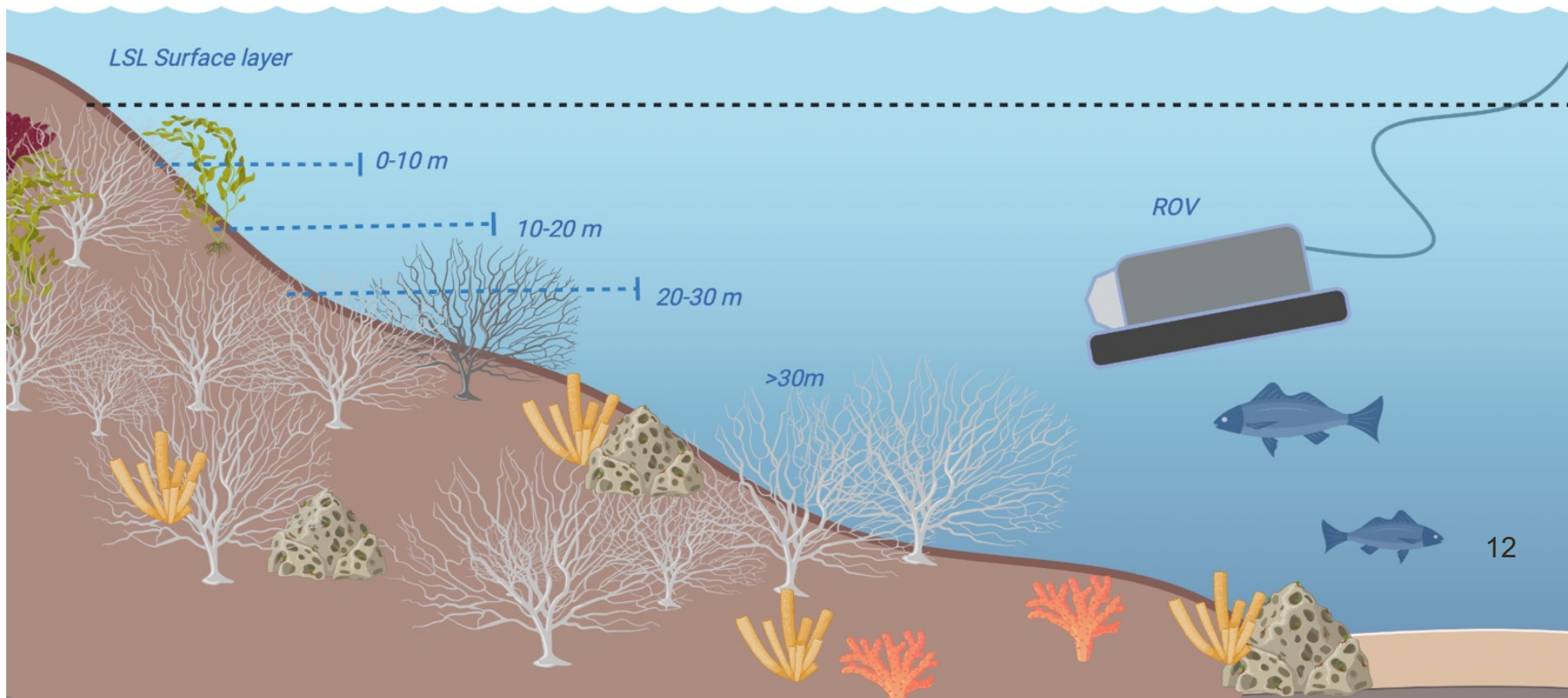
3 x 15 m x 2 m transects

15 m

ROV

3 x 15 m x 2 m transects

>50 m



## Data collection trips:

January 9<sup>th</sup>-14<sup>th</sup> 2023 – RV Southern Winds – focused on Doubtful and Thompson sounds

March 17<sup>th</sup>-22<sup>nd</sup> 2023 – MV Pembroke – focused on Dusky and Breaksea Sounds

May 17<sup>th</sup>-13<sup>th</sup> 2023 - MV Pembroke – focused on Dusky and Breaksea Sounds

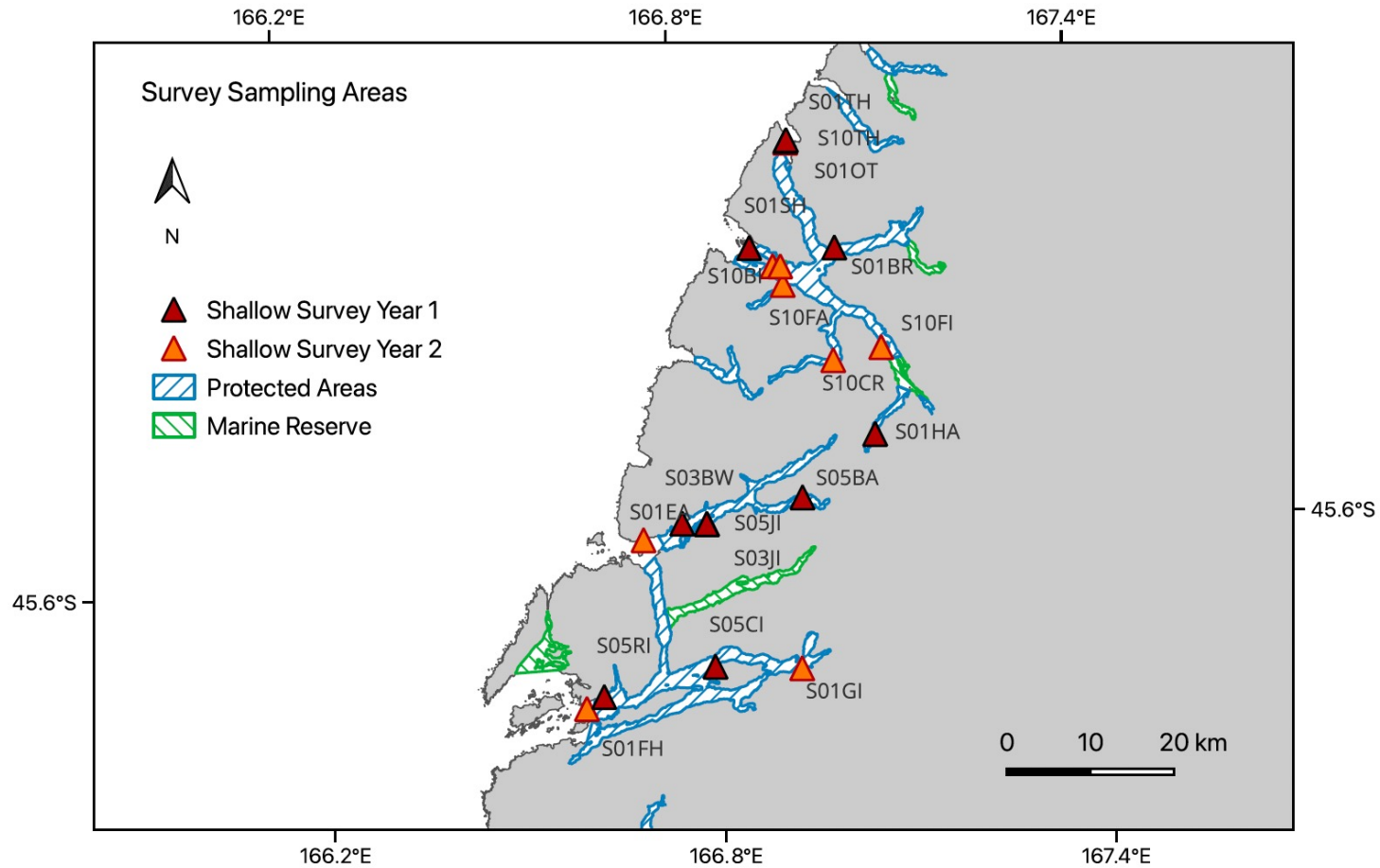
October 9<sup>th</sup>-14<sup>th</sup> 2023 - RV Southern Winds – focused on Doubtful Sound

January 8<sup>th</sup>-13<sup>th</sup> 2024 - RV Southern Winds – focused on Doubtful and Thompson sounds

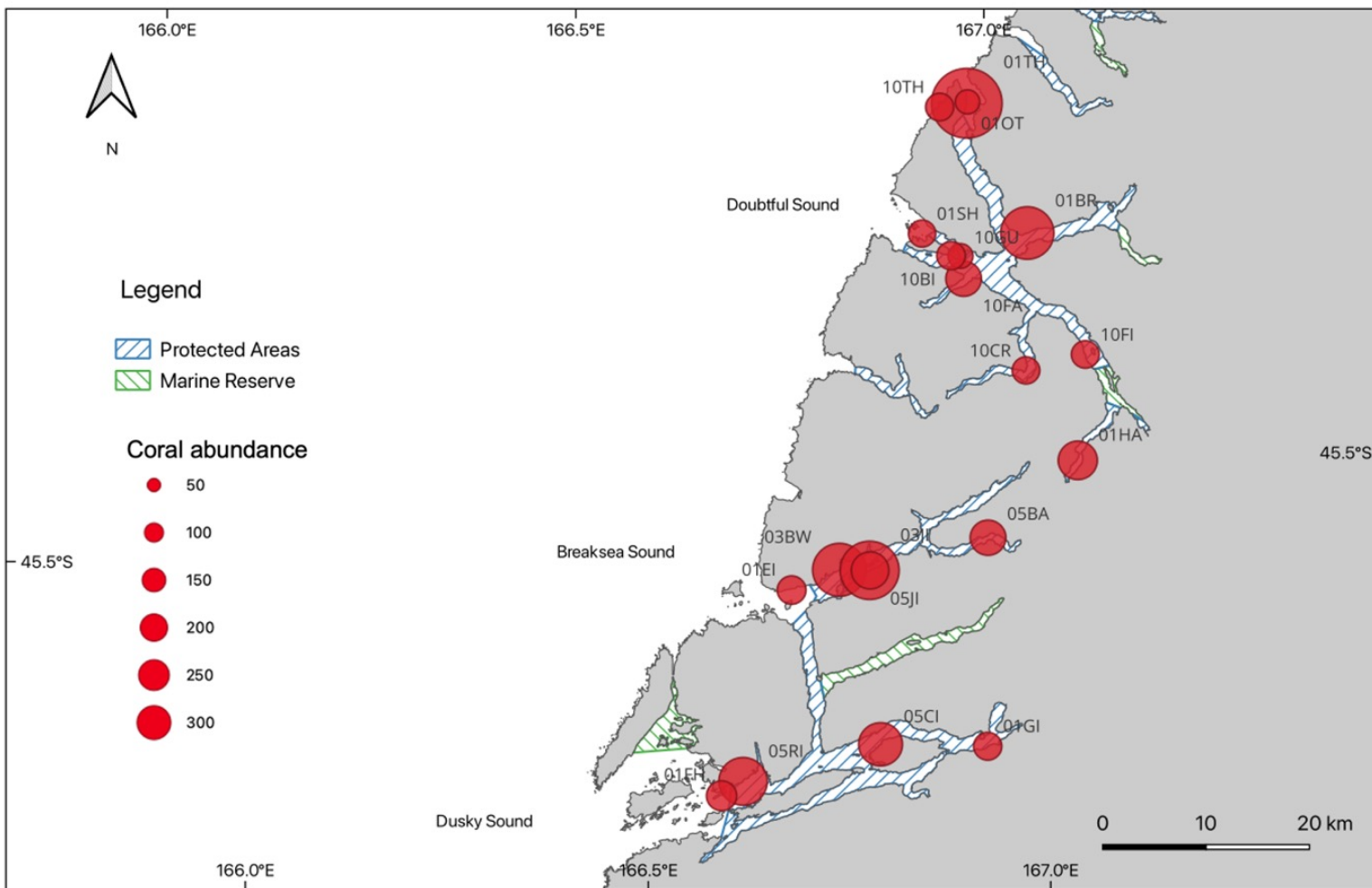
May 12<sup>th</sup>-18<sup>th</sup> 2024 - MV Pembroke – focused on Dusky and Breaksea Sounds (only included the collection of the remaining genetic samples, no other sampling)



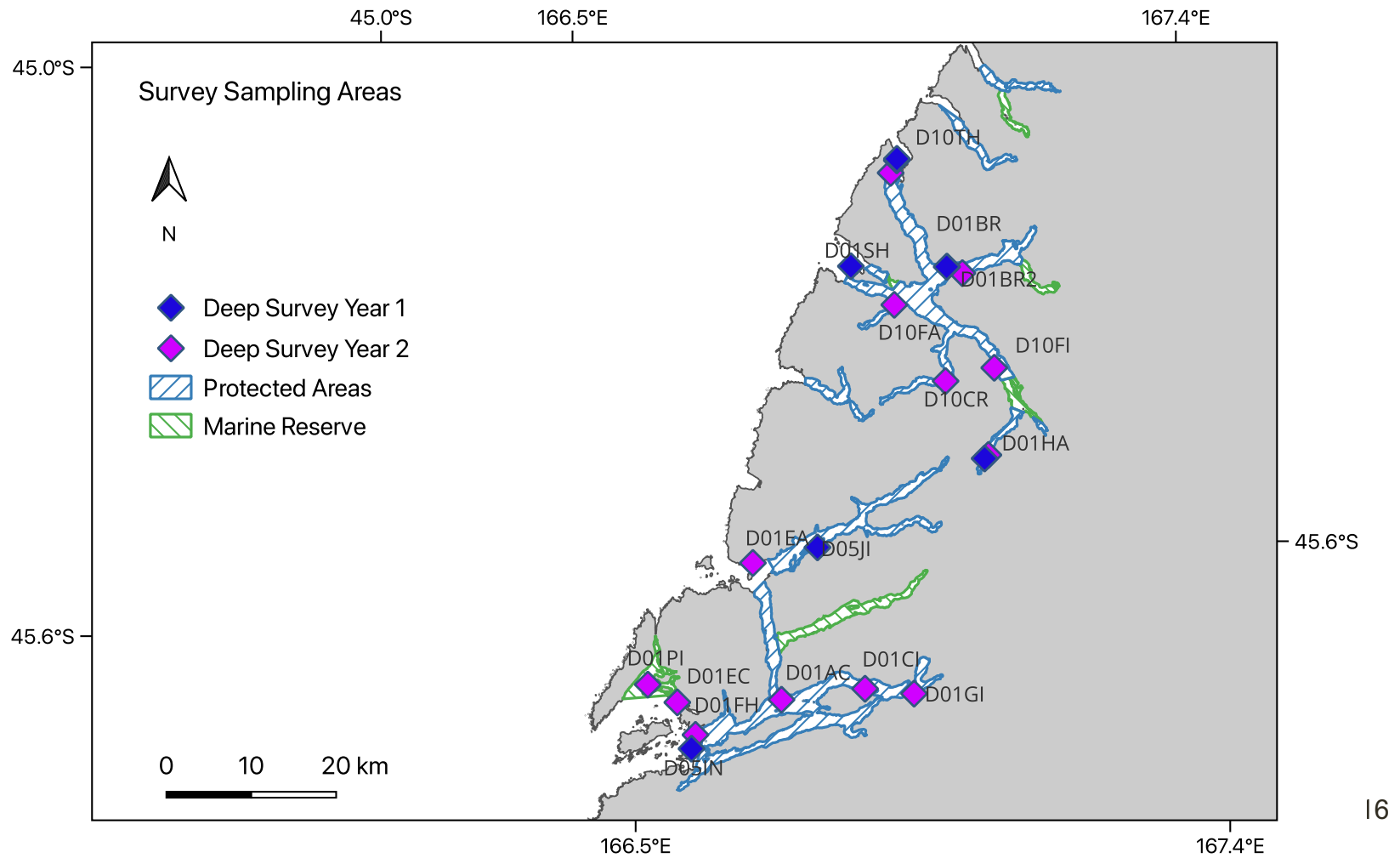
# Black Coral Abundance



Location of shallow black coral abundance surveys in Doubtful, Breaksea and Dusky Sounds



Total abundance of black corals (based on 3 x 15 x 2 m transects) across Doubtful, Breaksea and Dusky Sounds



Location of deep black coral abundance surveys in Doubtful, Breaksea and Dusky Sounds



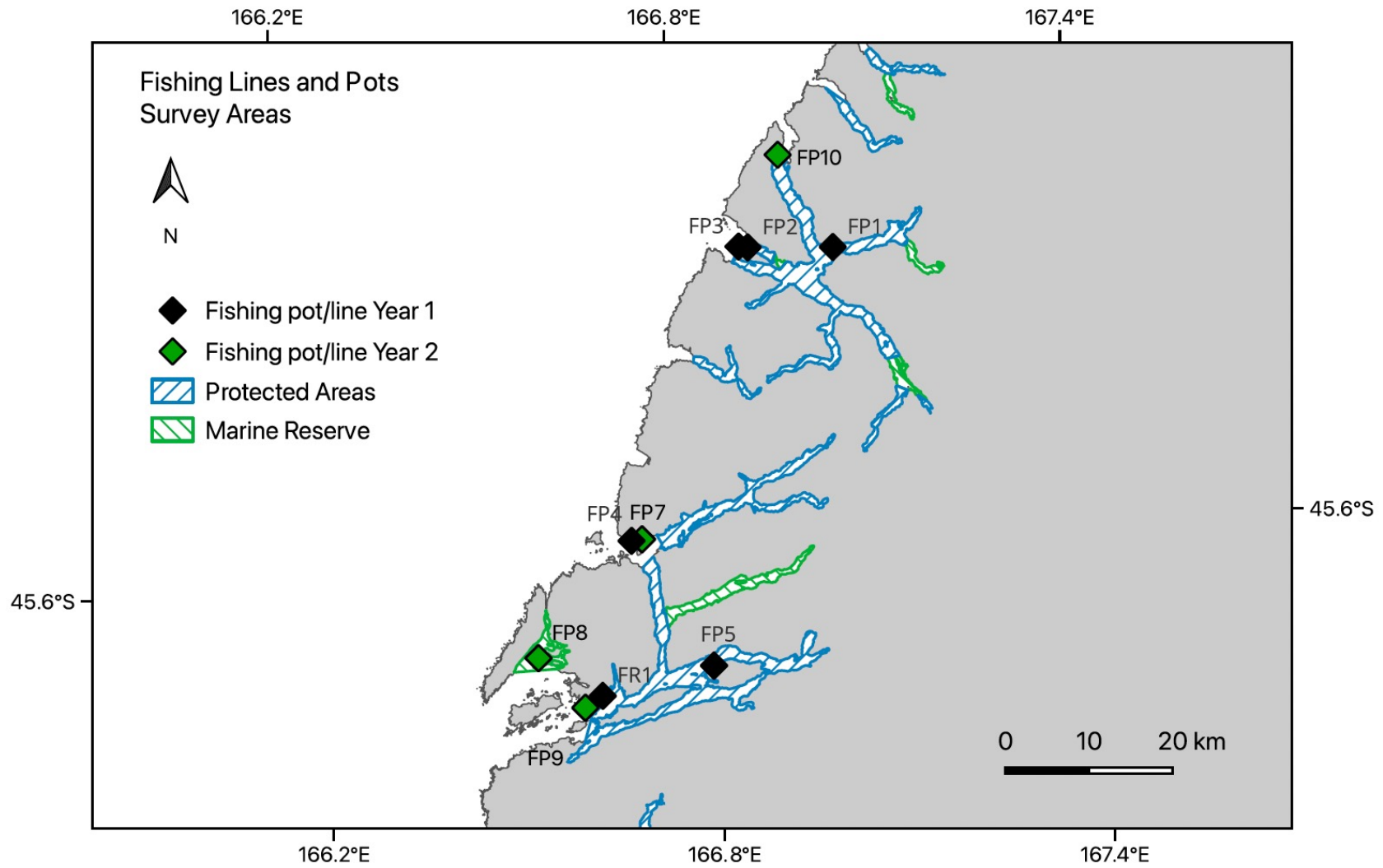
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# Approaches to this include:

- Document 'lost' pots and ropes
- Assess any damage to black coral populations
- Use previous catch databases and records to assess overlap between black coral and fisheries





Locations where 'lost' fishing pots have been found

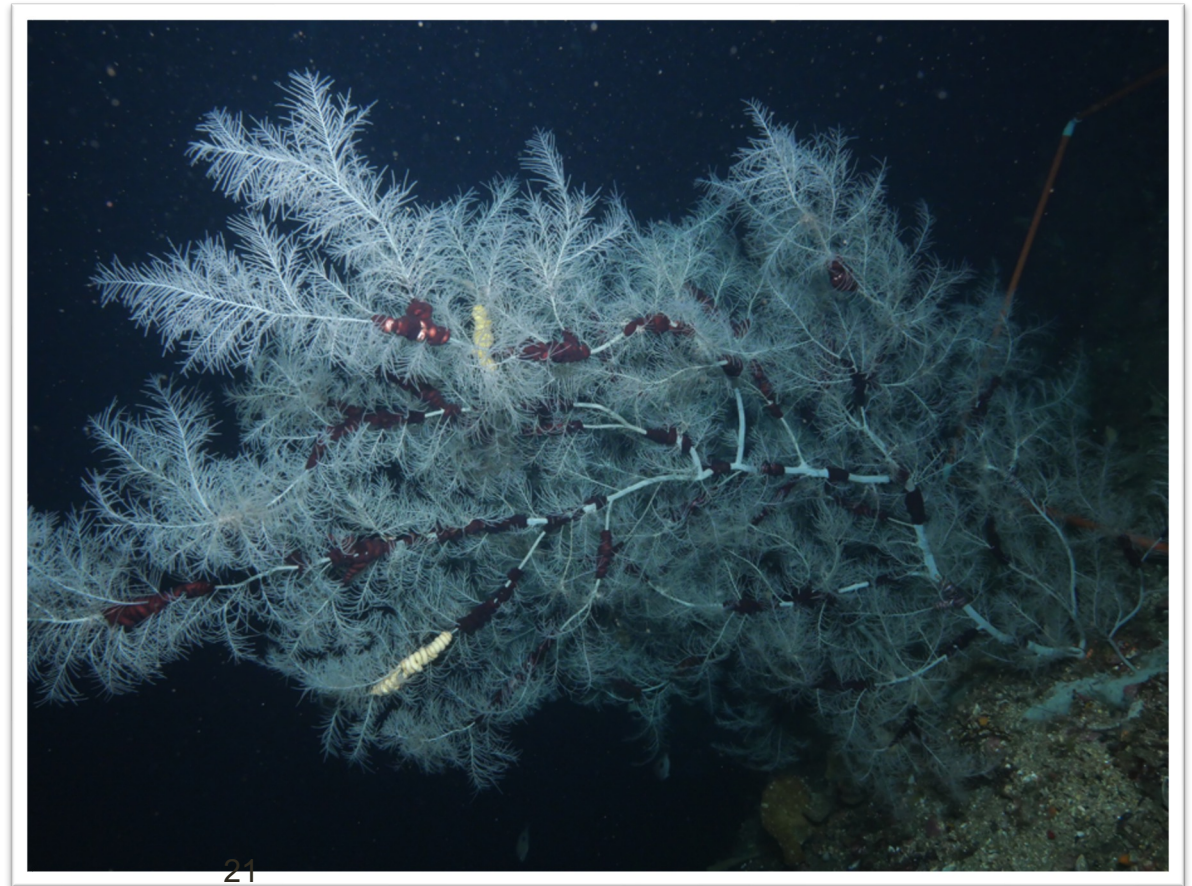


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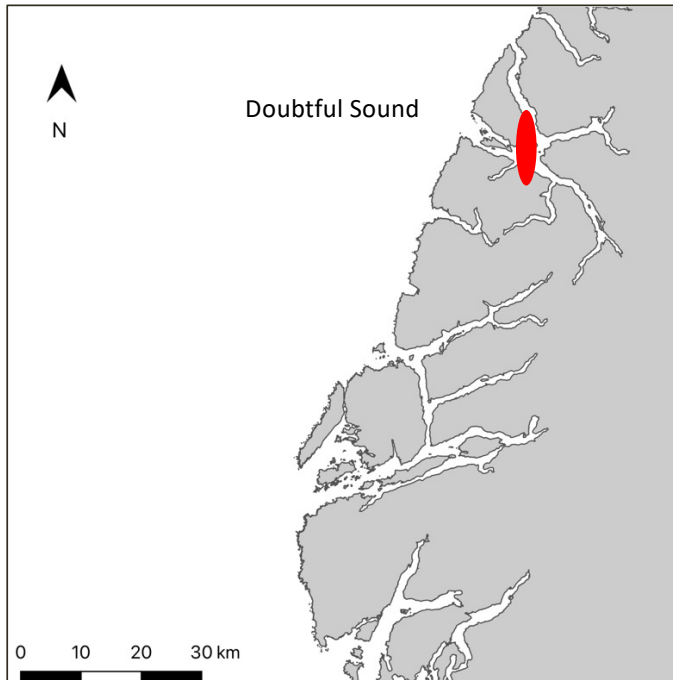
# Develop a population model to investigate how population may be impacted by changing environmental conditions

- Determine how demographic processes affect population dynamics and viability under different scenarios
- Predict the recovery of populations from environmental impacts



# Methods

## DATA SOURCING



## MODELING/STATISTICAL ANALYSIS

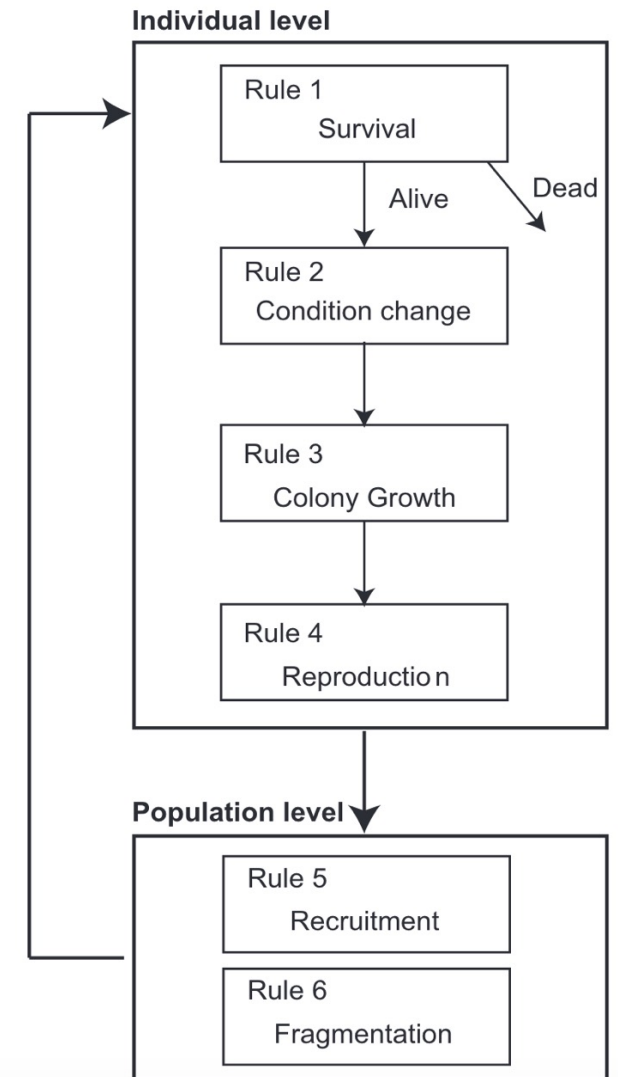
Size-age records from  
NIWA/Cawthron Institute  
dataset for Doubtful Sound



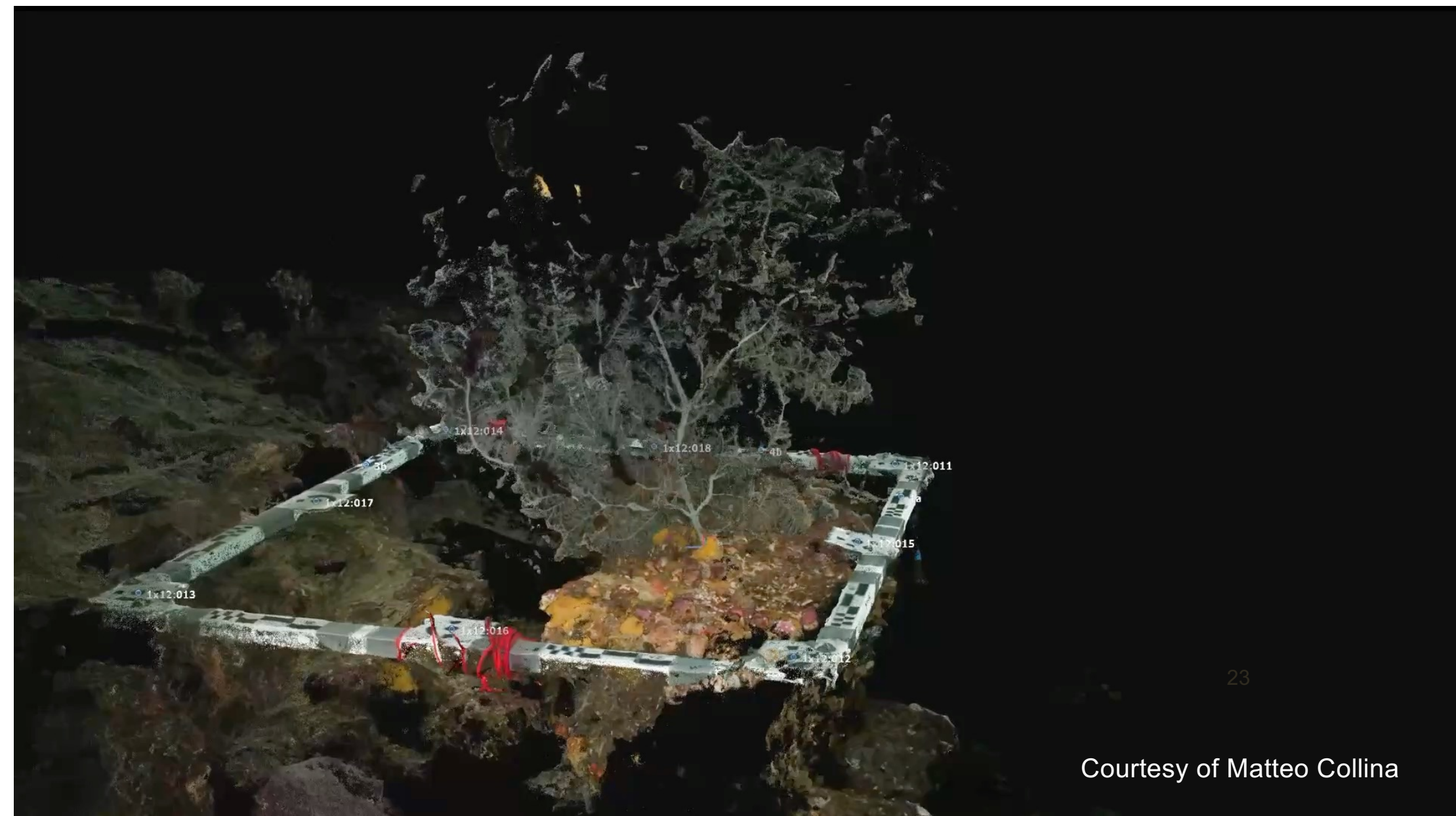
Report growth, recruitment  
and mortality



Simulate over time different  
scenarios with different  
recruitment rates





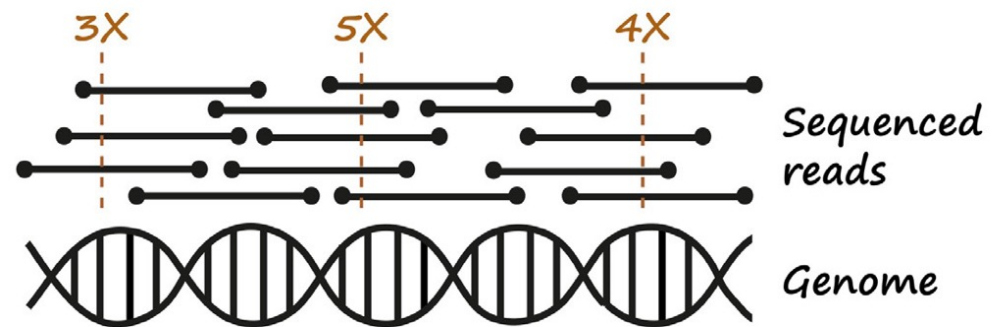
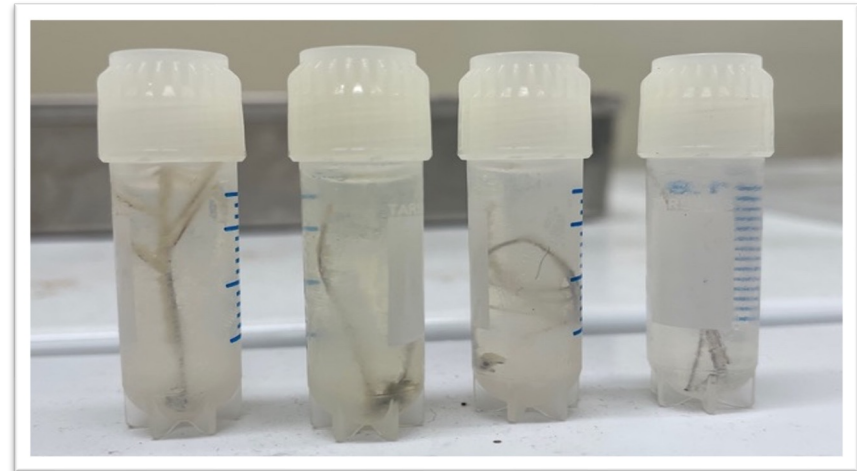


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## Assessing connectivity between populations using Genome-Wide SNPs

- Reconstruct a whole genome sequence to develop SNPs markers
- Assess whether populations are genetically distinct using SNPs markers
- Assess the extent of genetic connectivity across fiords and with depths



# Methods

## FIELD SAMPLING

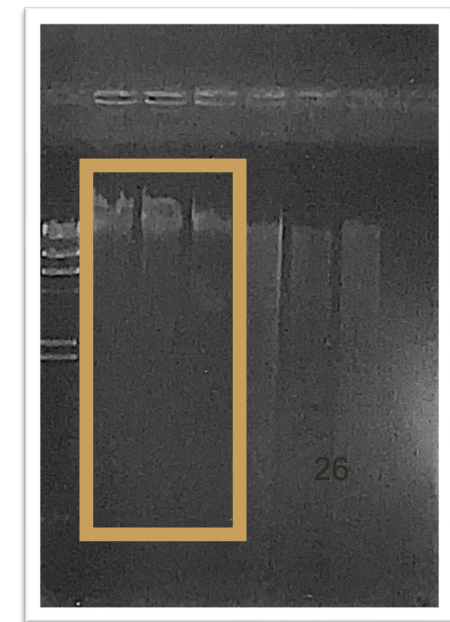
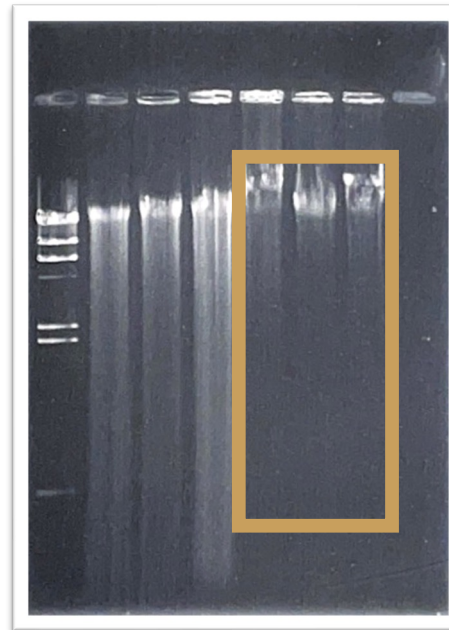
- DNA extraction
- Sequencing
- Genome assembly
- Library composition
- Population genetics

### 10 Sampling sites

8 shallow  
populations  
(0-20m)

2 shallow + deep  
populations  
(70-100m)

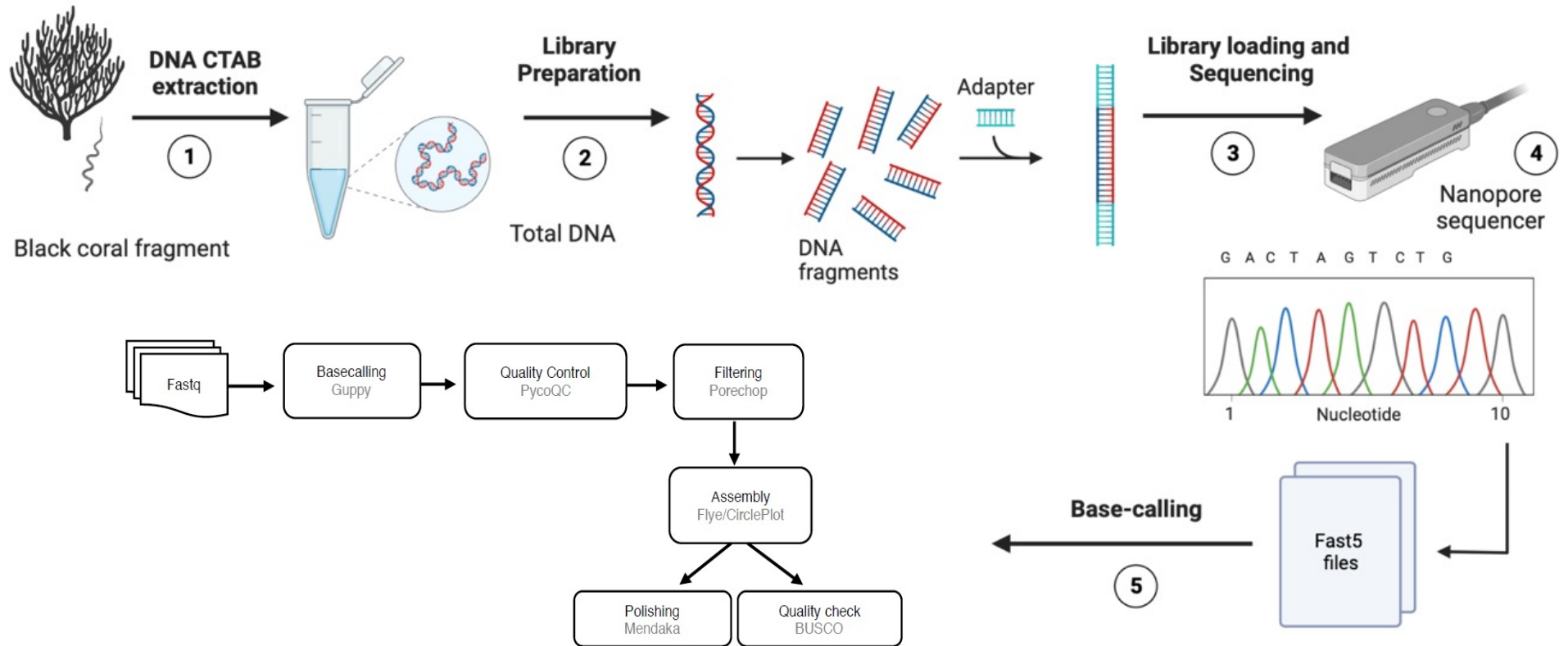
20-25  
individuals  
each





# Reference genome assembly workflow

## Methodology



# DNA extraction protocol

## Results



Trial 1 → Rapid Salt Extraction Protocol



Trial 2 → Qiagen DNeasy Blood & Tissue Kit



Trial 3 → CTAB (1%) extraction protocol



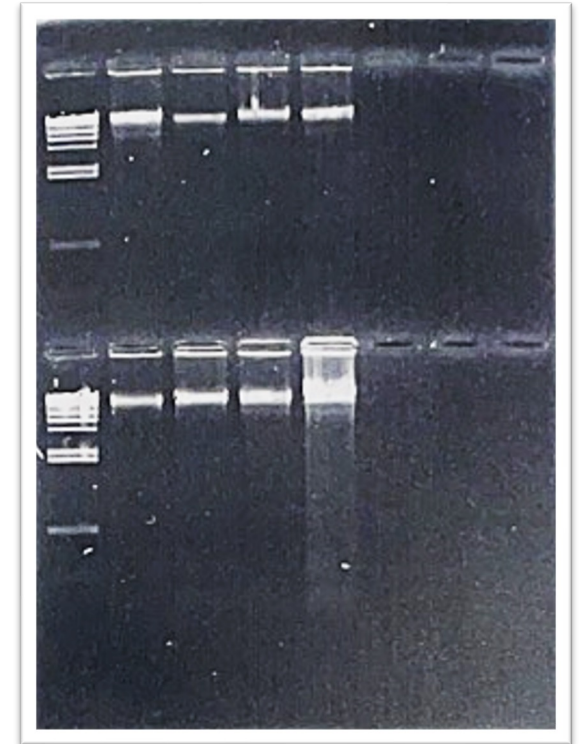
Trial 4 → CTAB(2%) 1 Phenol : Chloroform : Isoamyl  
(25:24:1) +

1 Chloroform : Isoamyl (24:1)



extractions

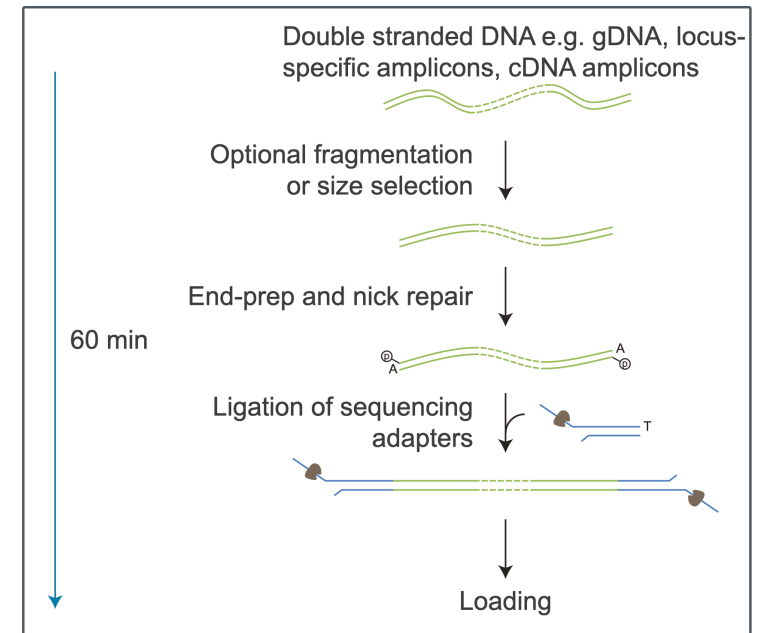
Trial 5 → CTAB (2%) 2 Chloroform: Isoamyl (24:1)  
extractions



# Library Preparation

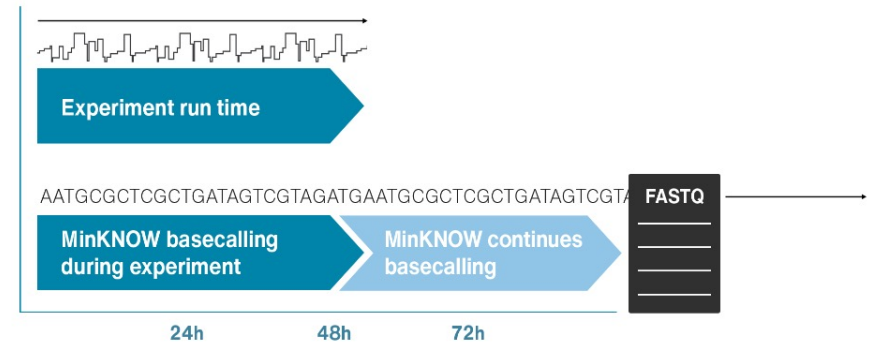
## Methodology

- Kit V14 SQK-LSK114
- Input 1  $\mu\text{g}$  of gDNA
- DNA repair and end-prep (Repair the DNA and prepare the DNA ends for adapter attachment)
- Adapter ligation and clean-up (attach the sequencing adapters to the DNA ends)



# Basecalling Methodology

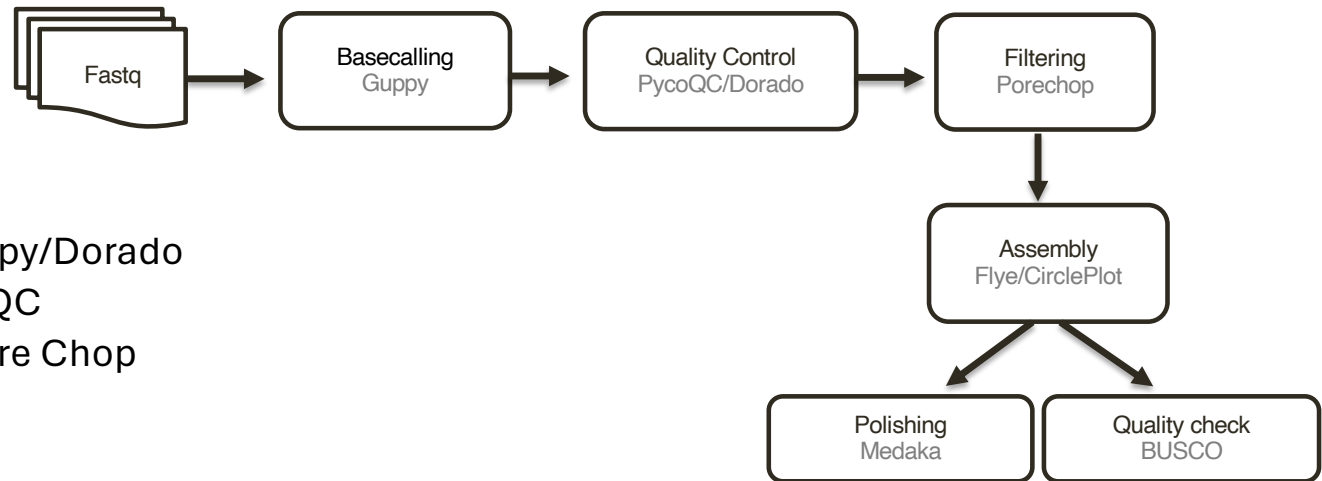
- The signal is stored in POD5 files
- Processed into FASTQ/BAM files
- Single PromethION gDNA run → 90Gb of sequence





# Bioinformatic pipeline

## Methodology

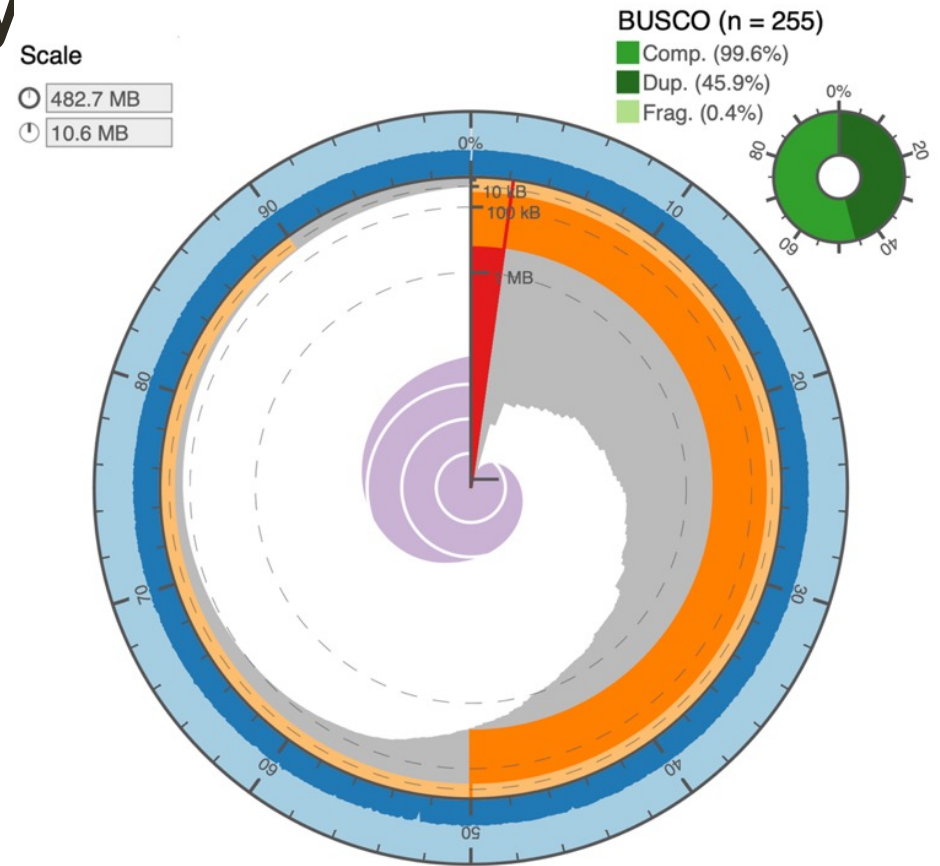


- Basecalled reads with Guppy/Dorado
- Quality control using PycoQC
- Filtering the reads using Pore Chop
- Assembled (Flye)
- Polishing
- Quality check

# Reference genome assembly

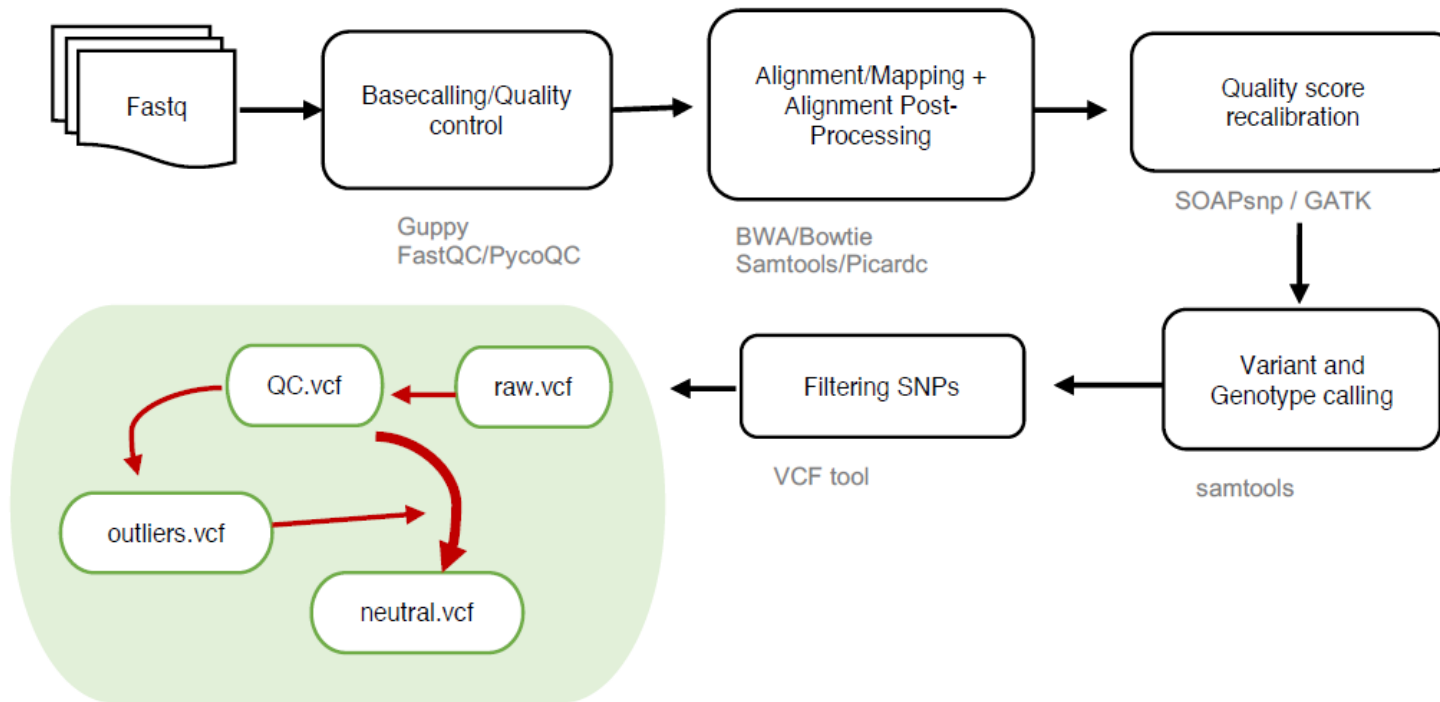
## Preliminary results

- WGS provides a comprehensive view of an organism's entire genome
- Detection of various types of genetic variations, including single nucleotide polymorphisms (SNPs), insertions, deletions, copy number variations (CNVs), and structural rearrangements
- allows for retrospective analysis and future investigations without the need for additional sequencing

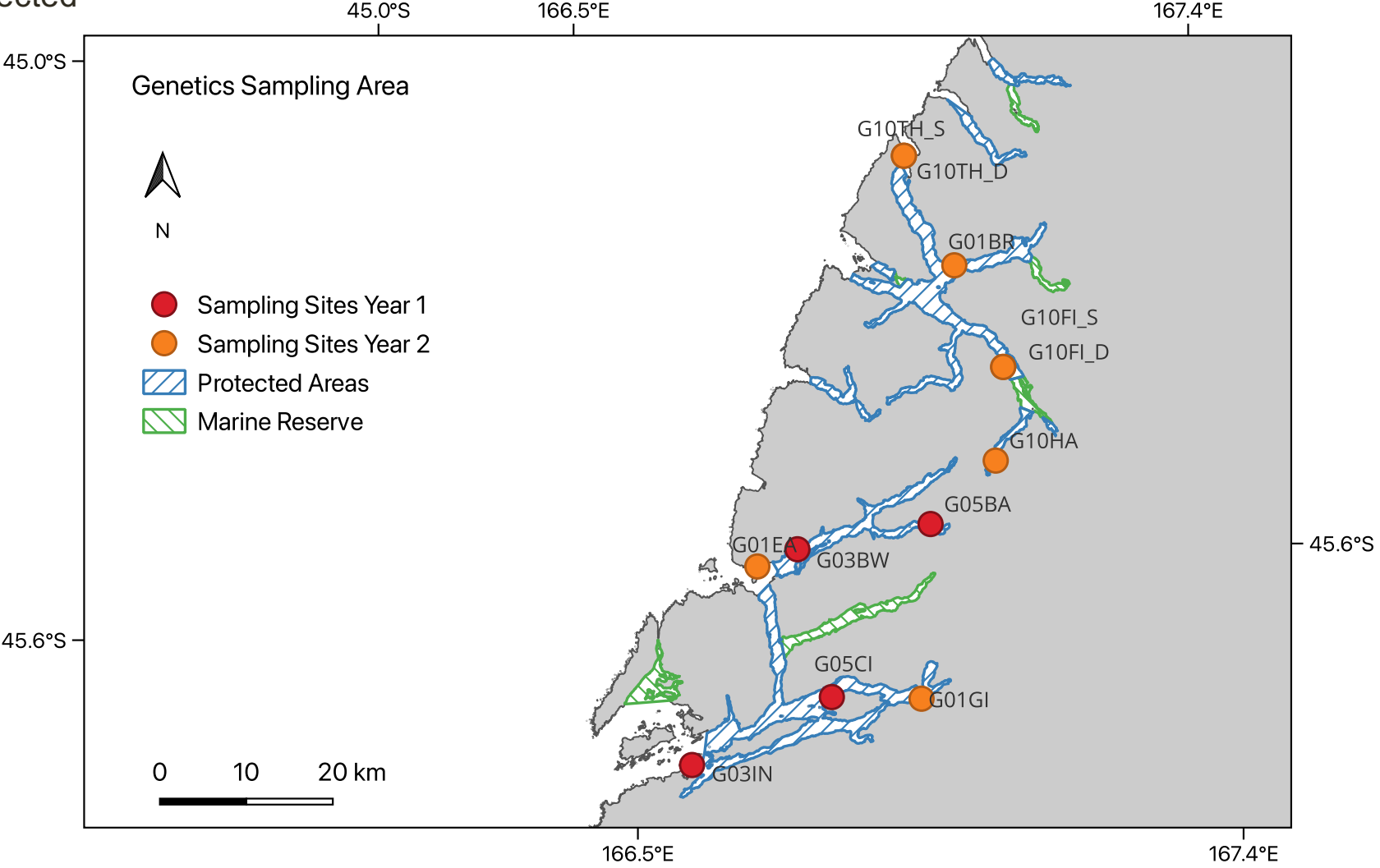


# Genetic population analysis pipeline

## Preliminary results



# Locations of genetic samples collected





Next steps....

# Acknowledgments

Bell research group  
Richard Kinsey and other crew/skippers on RV Southern Winds  
Dr Lyndsey Holland - Department of Conservation  
Southern Fiordland Initiative – Kathryn and Paul Mitchell  
Fiordland Marine Guardians – Rebecca McLeod



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